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MULTIPLE MATHEMATICAL CORRECTIONS BEFORE QUANTITATIVE ANALYSIS OF BIOLOGICAL INFRARED SPECTRA

Key words: Biological Mid-FTIR spectra, quantification, spectral correction.

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ABSTRACT

Infrared reflectance spectroscopy is often used to predict the composition of various food products. However the precision of the analysis could be improved. For this purpose, several treatments can be used ; (i) baseline correction. Baseline deformation is one of the major problem that is associated with infrared spectra; (ii) spectra can be expanded in terms of a set of orthonormal polynomials derived from the Legendre polynomials, and the leading terms of the expansion, which contains most of the baseline variation, can be removed; (iii) the classification of samples prior to analysis is one way to obtain more homogenous families. Hence, with the aim of improving the precision of the values obtained during quantitative determination of constituents from biological samples, we have successively applied different mathematical treatments on their infrared spectra : baseline correction, Legendre polynomials decomposition and classification procedure. The application of these mathematical treatments improves successively the precision of the predicted sucrose content values in biological samples. The mean and standard deviation of the differences between predicted and reference values are : 1.52×10^{-2} and 3.17×10^{-1} respectively before correction and (i) after the baseline correction : 5.06×10^{-2} and 3.08×10^{-1} % respectively, (ii) after decomposition Legendre polynomials : -6.13×10^{-3} and 2.76×10^{-1} % respectively, (iii) and after classification procedure according to correlation distance : -1.00×10^{-3} and 2.65×10^{-1} % respectively. Surprisingly, when only classification procedure is used for correction, the mean and SD values are 7.43×10^{-2} and 2.62×10^{-1} . Hence, it appears that the successive mathematical treatments can be advantageously substituted by a single classification procedure.

INTRODUCTION

Infrared reflectance spectroscopy is more and more used in food and agricultural industry for the quantitative determination of major biochemical constituents (Osborne and Fearn, 1986, Williams and Norris, 1987). With the recent developments in the field of computer and software engineering and the advent of new techniques such as ATR (Attenuated Total Reflectance) cells, the development and application of physical quantitative analysis methods such as Mid infrared spectroscopy have considerably expanded (Crocombe *et al.*, 1987; Van de Voort and Ismail, 1991). Principal Component Analysis showed to be very useful for the study of MIR spectra (Gillete and Koenig, 1982, Bertrand *et al.*, 1984, Cowe and Nocol., 1985).

We have previously reported the use of multidimensional statistical analysis combined with Mid-FTIR spectroscopy for the prediction of sucrose content sugar cane juices (Cadet *et al.*, 1991, Cadet and Offmann., 1995). However the precision of the analysis could be improved.

With the aim of improving the precision of the values obtained during quantitative determination of constituents from biological samples, we have directly or successively applied different mathematical treatments on their infrared spectra : baseline correction (Cadet and Offmann, 1996), decomposition Legendre polynomials (Cadet, 1996a) and classification procedure (Cadet, 1996b).

MATERIALS AND METHODS

Biological samples.

Raw sugar cane juices are obtained as previously described (Cadet and Offmann, 1995). The calibration set is constituted of 107 samples and the verification set of 30 samples.

Mid-FTIR ATR spectra

Mid-Fourier Transform Infrared (Mid-FTIR) spectra were collected on a Michelson-100 Fourier transform spectrophotometer. The data were recorded from 5000 to 500 cm^{-1} in 4 cm^{-1} increments at $\log(1/R)$, in which R is the ratio of the reflected intensity for the background to that of the sample. Attenuated total reflectance spectra were obtained with a Specac Overhead Attenuated Total Reflectance system. The crystal of the reflectance element is made from zinc selenide, a material that is quite inert to water; it is quite rapidly cleaned between samples by spraying with water and then dried with filter paper.

Mathematical treatment

All the mathematical treatments were performed on a Compaq personal computer with software written in "C" language and developed in our laboratory.

Multidimensional statistical analyses, such as principal component analyses (PCA), describe variation in multidimensional data by few synthetic variables. These synthetic variables are linear combination of all the original variables and have the advantage of having no correlation with each other. Simpler descriptions of data sets are thus obtained with minimal loss of information. These treatments were used for morphological analysis of spectra (le Nouvel., 1981) and for graphical representation of spectra similarity (Devaux *et al.*, 1988).

PCA was applied to the spectra from 800 to 1240 cm^{-1} (with 230 data points used as principal variables). Spectra were centered prior to PCA according to :

$$X_{ij} = A_{ij} - A_j - A_i + A$$

where X_{ij} = centered data ; A_{ij} = spectral data ($\log 1/R$) of spectrum i and wavelength j ; A_j = mean value of spectral data at wavelength j for every spectrum; A_i = mean value of spectral data of spectrum i for every wavelength; and A = average mean of all spectral data in the collection.

Principal component regression (PCR) was used to establish a prediction equation. PCR is basically a multilinear regression applied to scores assessed by PCA (Lebart *et al.*, 1977). Interest in the introduction of scores according to their predictive ability had already been shown (Bertrand *et al.*, 1987, Cadet & al., 1995).

Concentrations are predicted according to :

$$C_{n,l} = X_{n,k} \cdot V_{k,p} \cdot R_{p,l}$$

where C is the column vector of predicted concentrations, X is the centered matrix of spectral data, V is the matrix of latent vectors of PCA, and R is the column vector of the regression coefficients of the prediction equations. n , k , p are respectively the number of samples; the number of wavelengths; the number of significant principal components.

RESULTS AND DISCUSSION

Reference values and spectra.

The spectrum between 3800 and 700 cm^{-1} of a sample features three major absorption zones : 3700-2800 cm^{-1} , 1800-1470 cm^{-1} and 1240-800 cm^{-1} (Figure 1).

The first two zones can be assigned mainly to water. The 1240-800 cm^{-1} zone features the characteristic C-O, O-H and C-C vibrational bands of saccharides. In the spectrum, the

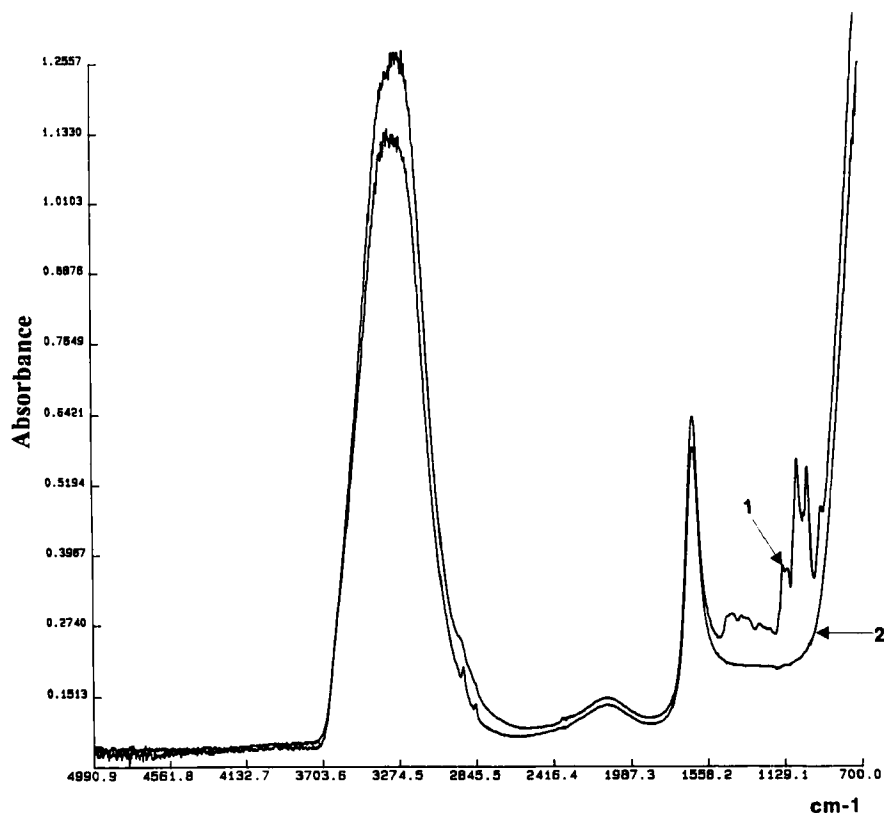


Figure 1. Mid-FTIR spectra of a biological sample (1) and of water (2).

1240-800 cm⁻¹ zone however corresponds to the absorption zones of the three major constituents of sugar cane juice (sucrose, glucose and fructose). The reference values associated with each of the spectrum of the verification set are given in Table 1.

Baseline correction

The deformation of baselines (Powell, 1984, Hruschka, 1987) is one of the problem that face infrared spectra. In MIR

Table 1. Difference between the reference and the predicted values without correction and after simple or successive corrections: baseline correction, decomposition Legendre polynomials, classification procedure.

SAMPLES NUMBER	Initial Spectrum		Mathematical Treatments										Direct classification	
	Reference (a)	Predicted (b)	Deviation (b-a)	Baseline correction (1)		(1) ± decomposition Legendre polynomials (2)		(1) ± (2) ± classification		(1) ± (2) ± classification		Class	Predicted (f)	Deviation (f-a)
				Predicted (c)	Deviation (c)	Predicted (d-a)	Deviation (d-a)	Predicted (e)	Deviation (e-a)	Predicted (e-a)	Deviation (e-a)			
1	21.029	21.185	0.166		21.201	0.172	21.108	0.078	2	21.027	-0.002	2	20.935	-0.094
2	22.144	22.082	-0.062		22.169	0.065	21.997	-0.147	2	21.98	-0.194	2	22.29	0.146
3	21.487	21.553	0.066		21.554	0.067	21.307	-0.19	2	21.086	-0.389	2	21.448	-0.041
4	21.899	21.649	-0.05		21.897	-0.002	21.788	0.098	2	21.913	0.214	2	21.755	0.056
5	23.335	23.419	0.084		23.313	0.278	22.999	0.264	2	22.986	-0.049	2	23.47	0.435
6	21.197	21.361	0.064		21.359	0.162	20.896	-0.301	2	21.136	-0.081	2	21.204	0.007
7	21.591	21.586	-0.005		21.572	0.278	21.84	0.259	2	22.591	0.68	2	22.034	0.453
8	24.273	24.028	-0.245		24.085	-0.187	24.151	-0.128	2	23.781	-0.482	2	24.672	0.399
9	19.753	20.48	0.637		20.316	0.563	20.325	0.532	2	20.185	-0.402	2	20.269	0.485
10	19.482	19.519	0.037		19.484	0.002	19.591	0.109	2	19.489	-0.013	4	19.63	0.148
11	21.967	21.135	-0.082		21.179	-0.009	20.997	-0.19	2	21.044	-0.143	2	20.827	-0.36
12	21.137	21.096	-0.131		20.951	-0.178	21.305	0.168	2	21.17	0.033	2	21.125	-0.012
13	20.171	20.026	-0.143		19.842	-0.229	19.972	-0.199	2	20.133	-0.038	2	19.889	-0.282
14	18.763	19.111	0.348		19.104	0.341	19.193	0.43	2	18.895	0.123	2	18.947	0.054
15	20.029	19.891	-0.098		20.146	0.117	20.058	0.029	2	19.98	-0.049	2	20.228	0.199
16	19.838	19.03	0.192		19.062	0.224	19.981	0.143	2	18.905	0.057	2	18.803	-0.095
17	19.373	19.787	0.414		19.537	0.164	19.497	0.124	2	19.58	0.187	2	19.489	0.116
18	21.017	21.37	0.353		21.211	0.194	21.489	0.472	2	21.256	0.239	2	21.259	0.242
19	20.804	21.019	0.215		20.841	0.037	20.642	0.038	2	20.872	0.068	2	21.132	0.319
20	20.884	20.478	-0.406		20.507	-0.376	20.709	-0.178	2	20.785	-0.179	2	20.895	0.328
21	20.16	20.41	0.25		20.11	-0.05	20.183	0.003	2	20.454	0.284	2	20.37	0.21
22	19.48	18.917	-0.563		18.98	-0.82	18.872	-0.608	2	18.984	-0.486	2	18.263	-0.187
23	20.223	19.824	-0.299		19.24	-0.283	19.276	-0.247	2	19.285	-0.333	2	20.261	0.271
24	20.281	20.434	0.153		20.509	0.473	20.389	0.268	2	20.27	-0.011	2	20.531	0.1
25	18.761	18.4	-0.361		19.354	-0.427	18.436	-0.348	2	19.541	0.059	2	19.48	-0.061
26	20.951	21.045	0.194		20.909	0.098	21.031	0.19	2	20.969	0.059	2	21.179	0.222
27	18.169	17.913	-0.256		17.789	-0.36	17.977	-0.182	2	18.327	0.158	2	18.193	0.023
28	17.04	16.338	-0.802		16.068	-0.972	16.491	-0.548	2	18.729	-0.317	2	18.41	-0.237
29	18.311	18.238	-0.073		18.188	-0.123	18.27	-0.041	2	18.401	0.09	2	18.568	0.237
30	17.548	17.648	0.06		17.335	-0.282	17.483	-0.108	2	17.446	0.355	2	17.787	0.199
Mean			1.82E-02			-8.98E-02		-4.13E-03			-1.94E-03			7.43E-02
Standard deviation			3.17E-01			2.98E-01		2.70E-01			2.61E-01			2.82E-01

spectra, uncertain variations are observed and these add an arbitrary constant to every absorbance. We have shown that baseline correction by total translation with respect to an invariable zone improves sensibly the precision of the quantitative predictions. We have extensively described this method previously (Cadet and Offmann, 1996).

From the spectra constitutive of the calibration set, the mean absorbance and the standard deviation for each and every point (wavelength) have been calculated. The diagram obtained (data not shown) features the zones at which the standard deviation (SD) value is close or equal to zero that is the invariable zones. Hence this zone has been chosen as the reference zone. The mean spectrum of the collected spectra has then been calculated and has been used as reference spectrum. Each of the other spectra of the collection undergo on the whole spectrum a translation (positive or negative) so that the spectra in the invariable zone is confounded.

The corrected spectra are obtained following :

$$S_c = S - cst$$

where S_c is the corrected spectra, S the initial spectra and cst the difference in absorbance between the invariable zone of the reference spectrum and the same zone in the spectrum to be corrected. The spectra of the calibration set and those of the verification set are thus corrected from the same reference spectrum. An example is given in figure 2.

The predicted values, without correction and with correction based on the invariable zone are given in Table 1. The precision of the predicted sucrose content values is improved for the corrected samples. For samples that have not been corrected, the bias and the standard deviation values are respectively $1.52 \cdot 10^{-2}$ and $3.17 \cdot 10^{-1} \%$ while for corrected samples (corrected from the invariable zone) the bias and SD

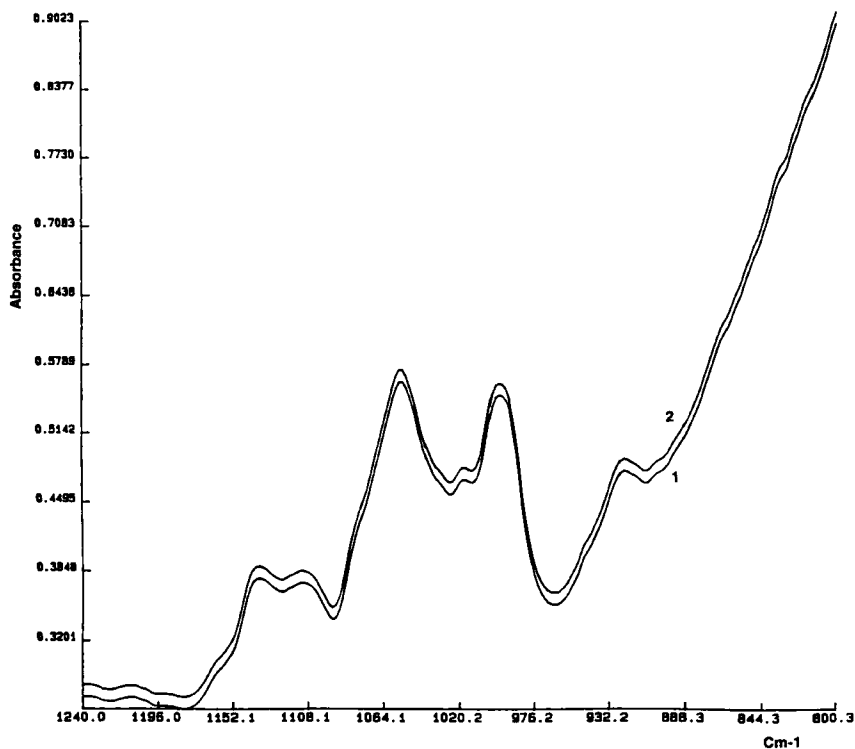


Figure 2. 800-1240 cm⁻¹ FTIR spectra before (1) and after (2) baseline correction (cst = 0.011).

are $-5.06 \cdot 10^{-2}$ and $3.08 \cdot 10^{-1} \%$ respectively. In fact SD values significantly decreases.

Decomposition by Legendre polynomials.

Spectra can be expanded in terms of a set of orthonormal polynomials derived from the Legendre polynomials, and the leading terms of the expansion, which contains most of the baseline variation, can be removed. The visual comparison of two spectra between α_1 cm⁻¹ and α_2 cm⁻¹ mainly consist in

comparing the areas beneath the spectra. A scalar product can hence be associated with the integral between α_1 , and α_2 .

Lipkus (1988) has introduced the use of an orthonormed polynomial Legendre system whose scalar product is given by :

$$\langle f, g \rangle = T \cdot \int_{\alpha_1}^{\alpha_2} f \cdot g \quad (1)$$

When the orthonormed system (α_1, α_2) is considered : ($P_i, i \in N$) a spectrum $S(\alpha)$ can be decomposed into :

$$S(\alpha) = \sum_{i=0}^{+\infty} \beta_i \cdot P_i \quad (2)$$

With the elimination of the first terms, it becomes :

$$S_0(\alpha) = S(\alpha) - \sum_{i=0}^n \beta_i \cdot P_i \quad (3)$$

where $S_0(\alpha)$ represents the corrected spectrum and n an integer.

We have recently described this method (Cadet, 1996a). The application of Legendre correction to the spectra for the filtering parameter n equal to 2 is illustrated on figure 3 as an example.

The mean of the difference between reference and predicted values are 1.52×10^{-2} before correction and $-6.13 \times 10^{-1} \%$ after correction, while standard deviation values are 3.17×10^{-1} and $2.76 \times 10^{-1} \%$ before and after correction (Table I). This treatment again improves the standard deviation value obtained after baseline correction.

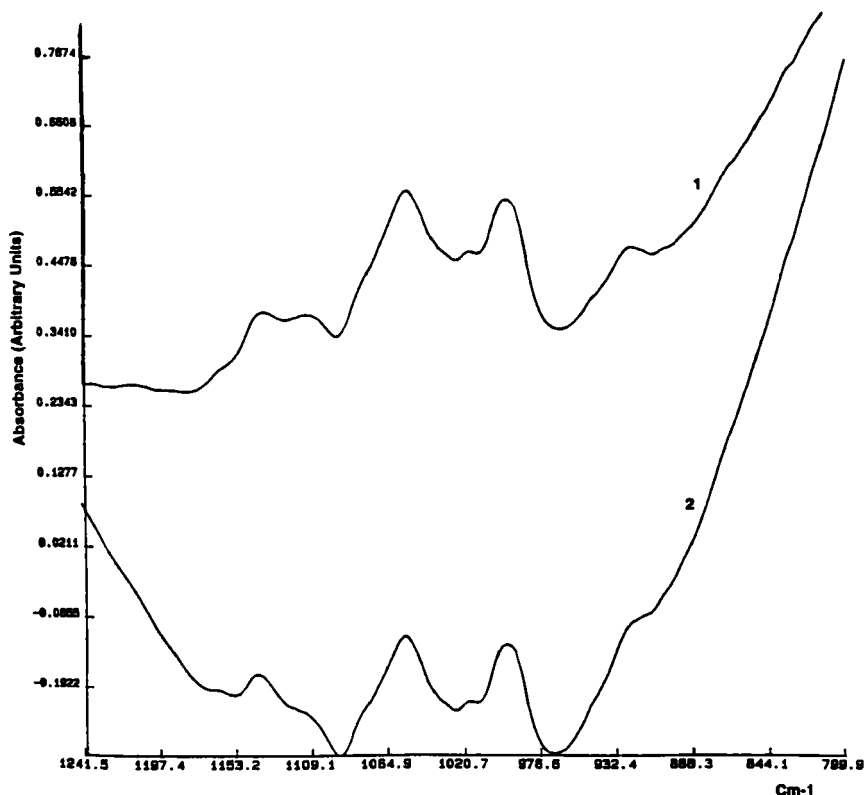


Figure 3. 800-1240 cm⁻¹ FTIR spectra before (1) and after (2) decomposition Legendre polynomials (n=2).

Classification procedure with the use of the correlation distance

The method extensively described in a recent paper (Cadet, 1996b) consisted in isolating the two most dissimilar individuals from a large calibration family of Mid-FTIR spectra and by successive Principal Component Analysis and Principal Component Regression, a family composed of a few individuals was constituted. Each individual from this family

represented the first spectrum of the corresponding classes that were ultimately formed. The classification of the remaining samples from the calibration family were carried out by the mobile centres method. The classification is done without knowing the chemical composition of samples.

Before the sucrose content value of a sample from the verification set is predicted, this sample is assigned to a class. The factorial map of the verification set (spectral collection) using the first two axes of the principal component analysis assessed on the calibration set is given in figure 4. We have shown that in a scope of quantifying sucrose in sugar cane juices the use of the euclidian distance in the classification procedure improves the results. However the correlation distance (half-defined and positive distance : the square of the sinus of two spectra) appears to be a more appropriate mathematical distance for the classification of Mid-infrared spectra of biological samples (Cadet, 1996c).

From the calibration family of 107 spectra, the four most dissimilar individuals were isolated and four classes were constituted. The classification procedure applied to each of these classes reduced the calibration family to 37 spectra that were distributed in 4 sub-classes. Each of the four sub-classes was assessed by principal component regression and four prediction equations were hence established. The mean and standard deviation of the differences between predicted and reference values were respectively 1.52×10^{-2} and $3.17 \times 10^{-1} \%$ before classification (one prediction equation is established with the whole calibration set) and -1.00×10^{-3} and $2.65 \times 10^{-1} \%$ after classification (Table 1). This procedure showed to improve, again, the precision of the predictions.

However, when the chemical values of the verification sets were directly predicted (without baseline correction and

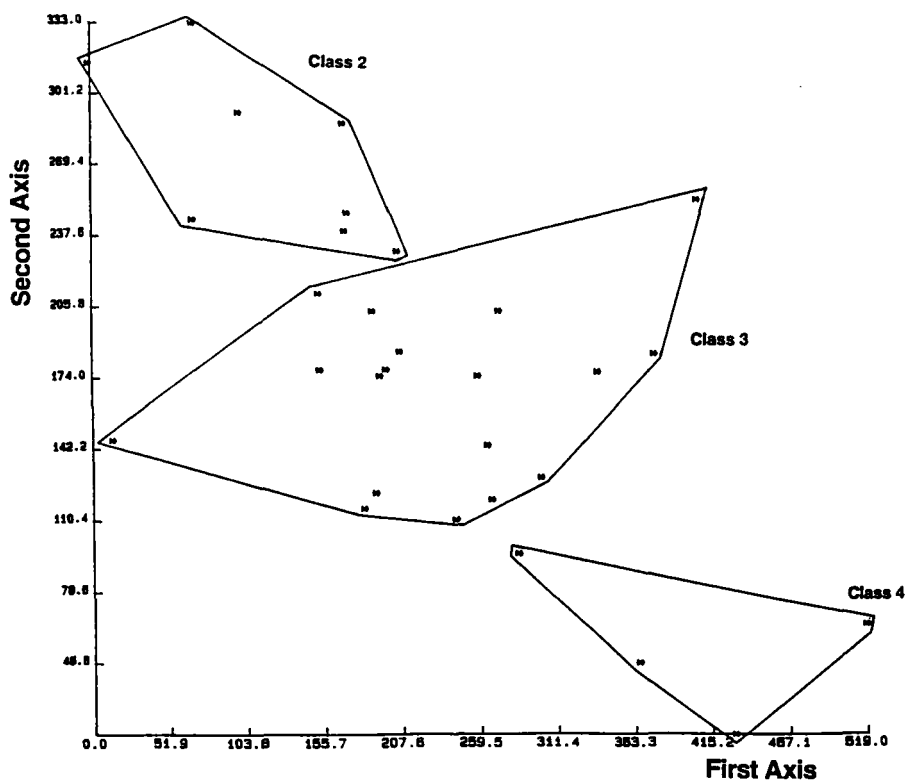


Figure 4. Classification of the verification set.

correction by Legendre polynomials decomposition) the mean and standard deviation were surprisingly 7.43×10^{-2} and 2.62×10^{-1} respectively.

CONCLUSION

The application of successive mathematical treatments to Mid-infrared spectra of biological samples (baseline correction, decomposition by Legendre polynomials and classification procedure) sensibly improves the precision of the predicted values of sucrose concentration.

The direct application of the classification procedure surprisingly gave similar results. It is known that the choice of the most appropriate mathematical distance in a classification procedure is difficult to make. We have recently shown that the correlation distance was more appropriate than the euclidian distance that is usually used (Cadet, 1996c). However the results obtained suggest that the mathematical distance used here (correlation distance) would not be the most appropriate mathematical distance. Even if the resulting classification procedure is not ideal, it appears to compensate aleatory translation errors (baseline correction) and baseline deformations (Legendre). This direct compensation is such that the result is sensibly better than that obtained after successive mathematical corrections.

These overall results show that the methods that have been proposed for the processing of Mid-IR spectra are interesting. The multiple mathematical treatment can be advantageously replaced by a single classification procedure hence simplifying considerably mathematical treatments applied to spectral data.

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